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Effect of cold helium plasma on the catalytic activity of certain erythrocyte dehydrogenases of rat blood

© Andrew K. Martusevich*, Anna G. Soloveva*, Svetlana Yu. Krasnova*,
Alexandr G. Galka*,**, Alexandr V. Kostrov**

* Prиволжский Research Federal University, Nizhny Novgorod, Russian Federation

** Institute of Applied Physics of RAS, Nizhny Novgorod, Russian Federation

Abstract: The work is aimed at clarifying the effect of cold helium plasma on the catalytic properties of lactate- and aldehyde dehydrogenase in rat blood erythrocytes. The effect was studied in 20 white Wistar rats. Upon completion of the full course of exposures (1 exposure per day for 5 days), blood samples were taken from all animals with subsequent erythrocyte isolation performed by standard differential centrifugation for assessing the activity of lactate dehydrogenase (LDH) and aldehyde dehydrogenase (AIDH). When assessing LDH activity, both direct and reverse reactions were considered. Gas flow microwave ionisation was applied to the synthesis of cold plasma using a special device developed at the Institute of Applied Physics, Russian Academy of Sciences. The plasma treatment was established to provide stimulation of LDH activity in both direct and reverse reactions. In the direct reaction, erythrocytic LDH activity in rats with plasma-treated skin almost doubled (94 %) against 48 % of activity growth in the reverse reaction. Rat blood erythrocyte AIDH tends to moderate inactivation, with catalytic properties observed to decrease by 13 %. Thus, treating the skin of healthy rats with cold helium plasma was demonstrated to stimulate the energy metabolism of blood cells, with moderate activity inhibition for AIDH presenting one of the detoxification enzymes. The nature of the observed shifts indicates their adaptability. In general, according to the obtained data, the modulation of free radical processes was confirmed to play a key role in the molecular-cellular mechanisms of the cold helium plasma action on the biological system.

Keywords: cold helium plasma, metabolic effects, lactate dehydrogenase, aldehyde dehydrogenase

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Влияние гелиевой холодной плазмы на каталитическую активность некоторых дегидрогеназ эритроцитов крови крыс

А.К. Мартусевич*, А.Г. Соловьева*, С.Ю. Краснова*, А.Г. Галка*,,
А.В. Костров****

* Приволжский исследовательский медицинский университет Минздрава России,
г. Нижний Новгород, Российская Федерация

** Федеральный исследовательский центр Институт прикладной физики РАН,
г. Нижний Новгород, Российская Федерация

Резюме: Цель работы – уточнение влияния гелиевой холодной плазмы на каталитические свойства лактатдегидрогеназы и альдегиддегидрогеназы эритроцитов крови крыс. На 20 белых крысах линии Wistar было изучено влияние гелиевой холодной плазмы на состояние эритроцитов. По

завершении полного курса воздействий (по 1 сеансу в течение 5 дней) у всех животных брали образцы крови и дифференциальным центрифугированием по стандартной методике выделяли из них взвесь эритроцитов для оценки активности дегидрогеназ – лактатдегидрогеназы в прямой и обратной реакциях, а также альдегиддегидрогеназы. Холодную плазму синтезировали с применением разработанного в Институте прикладной физики РАН специального устройства, работающего по принципу СВЧ-ионизации газового потока. Установлено, что изучаемый фактор обеспечивает стимуляцию активности лактатдегидрогеназы как в прямой, так и в обратной реакциях. У крыс, чьи кожные покровы были обработаны холодной плазмой, активность эритроцитарной лактатдегидрогеназы в прямой реакции практически удвоилась, продемонстрировав прирост в 94 %, а в обратной – повысилась лишь на 48 %. Альдегиддегидрогеназа эритроцитов крови крыс демонстрирует тенденцию к умеренной инактивации, что проявляется в снижении каталитических свойств на 13 %. Таким образом, показано, что обработка кожных покровов здоровых крыс гелиевой холодной плазмой обеспечивает стимуляцию энергетического обмена клеток крови, а также умеренно угнетает активность альдегиддегидрогеназы, одного из детоксикационных энзимов. Характер наблюдаемых сдвигов указывает на их адаптивность. В целом полученные в результате проведенного исследования данные подтверждают ведущую роль модуляции свободнорадикальных процессов в молекулярно-клеточных механизмах действия гелиевой холодной плазмы на биологическую систему.

Ключевые слова: гелиевая холодная плазма, метаболические эффекты, лактатдегидрогеназа, альдегиддегидрогеназа

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INTRODUCTION

In recent decades, the use of cold plasma in various therapeutic treatments has kindled the interest of biomedical specialists¹ [1–4]. The effectiveness of this treatment approach is confirmed by *in vitro* (on colonies of various microorganisms [1, 5, 6]) and *in vivo* (on infected skin abrasion models [5–7]) studies on the antibacterial activity of cold plasma-based therapies. The relevance of the antibacterial cold plasma effect is predetermined by the urgent need to find an alternative to antibiotics in the context of rapidly forming resistance in existing strains of pathogenic microorganisms [3, 5–7], though this has tended to eclipse research into other aspects of its effect on biological systems [1, 8, 9]. It should be noted here that both direct (at the place of treatment [4, 6, 9, 10]) and indirect (including systemic [1, 5]) realisation of these effects is possible. These particular aspects of the issue under discussion are discussed in less detail in the literature.

In previous *in vitro* experiments conducted by the authors, cold plasma was demonstrated to affect the oxidative and energy metabolism of biological systems along with their physical and chemical properties, as well as leading to an increase in antibacterial activity [8, 11–13]. Moreover, shifts were recorded in a number of metabolic parameters for blood [12] and the state of systemic haemodynamics and microcirculation in animals with dermal administration of the studied treatment [14, 15]. Although one possible cause of these transformations involves the effect on the catalytic activity of blood enzymes, no such confirmation of this hypothesis is provided in the experimental literature.

Against this background, the present study was aimed at clarifying the effect of cold helium plasma on the catalytic properties of lactate dehydrogenase (LDH) and aldehyde dehydrogenase (ALDH) in rat blood erythrocytes.

EXPERIMENTAL PART

The study was performed on 20 healthy sexually-mature male Wistar rats. The animals were divided into two groups equal in number. The first group ($n = 10$) presented a control group with no manipulations performed other than a single blood sampling. Rats of the second group ($n = 10$) were treated daily over the course of 5 days with cold helium plasma applied to a pre-epilated dorsal surface. A single treatment time comprised 1 min. Cold plasma was synthesised using a special device developed at the Institute of Applied Physics, Russian Academy of Sciences and based on the phenomenon of gas flow microwave ionisation [8, 11–15]. Grade A bottled helium was used as a source gas for obtaining the cold plasma.

¹ Aleinik A.N. Plasma medicine: textbook. Tomsk: Publishing house of Tomsk Polytechnic University, 2011. 45 pp.

Upon completion of the full course of exposure to cold helium plasma, blood samples were taken from all animals. Subsequent isolation of an erythrocyte suspension was carried out by differential centrifugation according to the standard method for evaluating the activity of lactate dehydrogenases (LDH) and aldehyde dehydrogenase (ALDH). The LDH activity was determined in the erythrocyte hemolysate admixed with distilled water (1:40 vol.) according to the method of G.A. Kochetov². Both direct and reverse reactions were considered for evaluation of LDH activity.

ALDH activity was determined spectrophotometrically according to the method described by B.M. Kershengolts and E.V. Serkina (1981). The protein content was specified using the modified Lowry method.

Statistical data processing was carried out using Microsoft Excel 2007 and the Primer of Biostatistics 4.03 software program.

RESULTS AND DISCUSSION

Cold helium plasma was shown to contribute to significant changes in the catalytic properties of erythrocyte enzymes. The studied treatment was observed to stimulate LDH activity in both direct and reverse reactions (see Fig. 1). However, the extent of these shifts is not the same: at the end of the course of exposure to cold plasma, the catalytic enzyme activity in both reactions is practically equalised, while, in the intact animals of the first group, a moderate predominance of the reverse reaction occurs. In this regard, in rats treated with cold plasma, the activity of erythrocyte LDH in the direct reaction almost doubled (94 %, $p < 0.05$ as compared to the intact animal level). Conversely, in the reverse reaction an increase of 48 % ($p < 0.05$) was observed. Indirectly, this can be seen as indicating a stimulating effect of the considered treatment on the intermediate element of energy metabolism with an increase in the production of pyruvate, the primary substrate of the Krebs cycle [16, 17].

A different aspect of the modification was revealed with respect to ALDH (see Fig. 2). The specified enzyme related to the enzyme detoxification system tends to a moderate inactivation with catalytic properties decreased by 13 % ($p < 0.05$ relative to the level detected for intact rats of the first group). This may be due to increased production of free radicals induced by external exposure to cold plasma. Although these compounds are intensively utilised by the antioxidant system of blood and tissues, their secondary (malon-dialdehyde) and tertiary (Schiff's bases) products require the involvement of appropriate enzymes in the detoxification process [18].

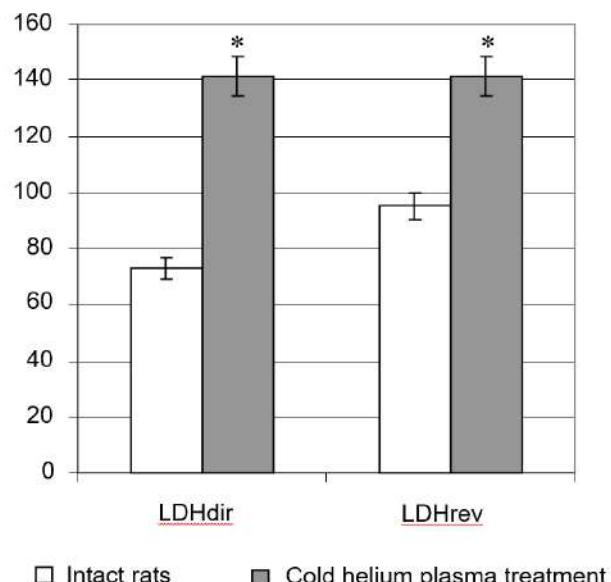


Fig. 1. Lactate dehydrogenase activity in direct (LDHdir) and reverse (LDHrev) reactions in rat blood erythrocytes (nM NADH/min per mg protein; * – statistical significance of differences relative to the intact animal level)

Рис. 1. Активность лактатдегидрогеназы в прямой (ЛГПр) и обратной (ЛДГОбр) реакциях в эритроцитах крови крыс (в нмольД НАДН/мин*мг белка; * – статистическая значимость различий по отношению к уровню, характерному для интактных животных)

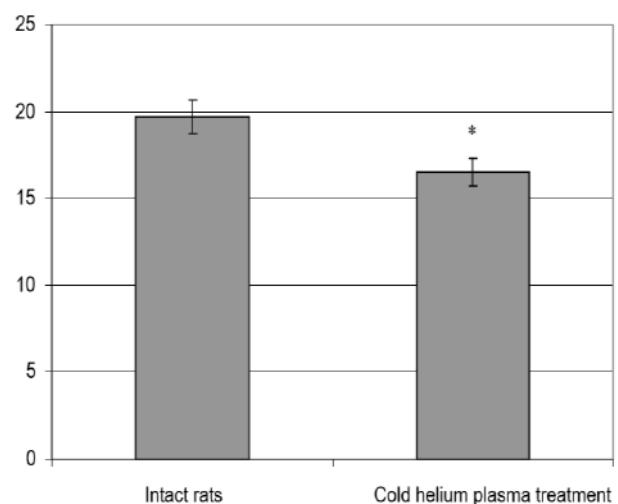


Fig. 2. Aldehyde dehydrogenase activity in rat blood erythrocytes (nM NADH/min per mg protein; * – statistical significance of differences in relation to the intact animal level)

Рис. 2. Активность альдегиддегидрогеназы эритроцитов крови крыс (в нмоль НАДН/мин*мг белка; * – статистическая значимость различий по отношению к уровню, характерному для интактных животных)

² Kochetov G.A. Enzymology practical guide: textbook for students of biological specialities. Moscow: Vysshaya Shkola, 1980. 272 pp.

This is confirmed by the increase in the concentration of malondialdehyde in rat blood erythrocytes exposed to the cold helium plasma demonstrated by the authors in previous studies [12, 13]. Moreover, no stimulating effect of cold plasma on the intensity of free radical reactions in the membranes of these blood cells was registered. For this reason, together with the results obtained in the framework of this study, the plasma effect can be characterised as training and pro-adaptive.

It is also important to emphasise that the presented data indirectly indicate the consistency of the working hypothesis on the induction of free radical processes as one of the main systemic effects caused by cold helium plasma to the body, since all detected metabolic transformations resulted from the studied treatment are directly associated with a short-term

increase in the intensity of free radical oxidation in the blood and tissues.

CONCLUSION

Thus, according to the dynamics of the catalytic properties of erythrocyte lactate dehydrogenase in direct and reverse reactions, treating the skin of healthy rats with cold helium plasma was established to stimulate the energy exchange of blood cells with simultaneous moderate inhibiting AIDH activity – one of the detoxification enzymes. The nature of the observed shifts indicates their adaptability. In general, according to the obtained data, the modulation of free radical processes was confirmed to play a key role in the molecular-cellular mechanisms of the cold helium plasma action on the biological system.

REFERENCES

- 1.** Dobrynin D, Fridman G, Friedman G, Friedman AA. Physical and biological mechanisms of direct plasma interaction with living tissue. *New Journal of Physics*. 2009;11(11):115020–115046. <https://doi.org/10.1088/1367-2630/11/11/115020>
- 2.** Dubuc A, Monsarrat P, Virard F, Merbahi N, Sarrette JP, Laurencin S, et al. Use of cold-atmospheric plasma in oncology: a concise systematic review. *Therapeutic Advances in Medical Oncology*. 2018;10. 12 p. <https://doi.org/10.1177/1758835918786475>
- 3.** Hoffmann C, Berganza C, Zhang J. Cold Atmospheric Plasma: methods of production and application in dentistry and oncology. *Medical Gas Research*. 2013;3(1):21. <https://doi.org/10.1186/2045-9912-3-21>
- 4.** Jawaid P, Rehman MU, Zhao QL, Takeda K, Ishikawa K, Hori M, et al. Helium-based cold atmospheric plasma-induced reactive oxygen species-mediated apoptotic pathway attenuated by platinum nanoparticles. *Journal of Cellular and Molecular Medicine*. 2016;20(9):1737–1748. <https://doi.org/10.1111/jcmm.12880>
- 5.** Ermolaeva SA, Varfolomeev AF, Chernukha MYu, Yurov DS, Vasiliev MM, Kaminskaya AA, et al. Bactericidal effects of non-thermal argon plasma in vitro, in biofilms and in the animal model of infected wounds. *Journal of Medical Microbiology*. 2011; 60(1):75–83. <https://doi.org/10.1099/jmm.0.020263-0>
- 6.** Alkawareek MY, Gorman SP, Graham WG, Gilmore BF. Potential cellular targets and antibacterial efficacy of atmospheric pressure non-thermal plasma. *International Journal of Antimicrobial Agents*. 2014;43(2):154–160. <https://doi.org/10.1016/j.ijantimicag.2013.08.022>
- 7.** Flynn PB, Busetti A, Wielogorska E, Chevalier OP, Elliott CT, Laverty G, et al. Non-thermal Plasma Exposure Rapidly Attenuates Bacterial AHL-Dependent Quorum Sensing and Virulence. *Scientific Reports*. 2016;6. Article number: 26320. <https://doi.org/10.1038/srep26320>
- 8.** Martusevich AK, Soloveva AG, Krasnova SYu, Yanin DV, Galka AG, Kostrov AV. The influence of helium cold plasma on metabolic and physical-chemical parameters of human blood in vitro. *Biomeditina = Journal Biomed*. 2018;2:47–58. (In Russian)
- 9.** Brun P, Pathak S, Castagliuolo I, Palù G, Zuin M, Cavazzana R, et al. Helium generated cold plasma finely regulates activation of human fibroblast-like primary cells. *PLoS ONE*. 2014;9(8):e104397. <https://doi.org/10.1371/journal.pone.0104397>
- 10.** Wiegand C, Fink S, Beier O, Horn K, Pfuch A, Schimanski A, et al. Dose- and time-dependent cellular effects of cold atmospheric pressure plasma evaluated in 3D skin models. *Skin pharmacology and physiology*. 2016;29(5):257–265. <https://doi.org/10.1159/000450889>
- 11.** Martusevich AK, Soloveva AG, Yanin DV, Galka AG, Krasnova SYu. The influence of helium cold plasma on the parameters of blood oxidative metabolism in vitro. *Vestnik novykh meditsinskikh tekhnologii = Journal of New Medical Technologies*. 2017;24(3):163–166. (In Russian) https://doi.org/10.12737/article_59c4a9e679ca86.74880803
- 12.** Martusevich AK, Soloveva AG, Krasnova SYu. Influence of helium cold plasma on the state of oxidative metabolism of rat blood. *Vestnik Ul'yanovskoi gosudarstvennoi sel'skohozyaistvennoi akademii = Vestnik of Ulyanovsk State Agricultural Academy*. 2018;2:161–165. (In Russian) <https://doi.org/10.18286/1816-4501-2018-2-161-165>
- 13.** Martusevich AK, Soloveva AG, Galka AG, Kozlova LA, Yanin DV. The effect of helium cold plasma on red blood cell metabolism. *Bulleten' eksperimental'noi biologii i meditsiny = Bulletin of Experimental Biology and Medicine*. 2019;167(2):144–146. (In Russian)
- 14.** Martusevich AK, Krasnova SYu, Peretyagin PV, Galka AG, Golygina ES, Kostrov AV. The Influence of Helium-Generated Cold Plasma on Parameters of Heart Rate Variability in Rats. *Bio-*

fizika = Biophysics. 2019;64(3):596–600. (In Russian) <https://doi.org/10.1134/S0006302919030219>

15. Martusevich AK, Krasnova SYu, Galka AG, Peretyagin PV, Yanin DV, Kostrov AV. Estimation of Microcirculatory Response to the Influence of Cold Helium Plasma. *Biofizika* = Biophysics. 2019;64(4): 767–771. (In Russian) <https://doi.org/10.1134/S0006302919040161>

16. Martusevich AK, Soloveva AG, Peretyagin SP. Influence of free and bound nitric oxide on properties of erythrocytes aldehyde dehydrogenase. *Voprosy biologicheskoi, meditsinskoi i farmatsevticheskoi khimii* = Problems of biological, medical and pharmaceutical chemistry. 2014;11:

60–65. (In Russian)

17. Davydov AV, Martusevich AK, Soloveva AG, Karimova RG. Metabolic adaptation of oxidoreductases of erythrocytes to glutathione-containing di-nitrosoyl iron complexes. *Uchenye zapiski Kazanskoi gosudarstvennoi akademii veterinarnoi meditsiny im. N.E. Baumana* = Bulletin of Kazan State Academy of Veterinary Medicine named after N.E. Baumana. 2015;221(1):60–64. (In Russian)

18. Soloveva AG, Peretyagin SP. The effect of subchronic inhalations of nitric oxide on metabolic processes in blood of experimental animals. *Biomeditsinskaya khimiya*. 2016;62(2):212–214. (In Russian) <https://doi.org/10.18097/PBMC20166202212>

БИБЛИОГРАФИЧЕСКИЙ СПИСОК

1. Dobrynin D., Fridman G., Friedman G., Friedman A.A. Physical and biological mechanisms of direct plasma interaction with living tissue // New Journal of Physics. 2009. Vol. 11. Issue 11. P. 115020–115046. <https://doi.org/10.1088/1367-2630/11/11/115020>

2. Dubuc A., Monsarrat P., Virard F., Merbahi N., Sarrette J.P., Laurencin S., et al. Use of cold-atmospheric plasma in oncology: a concise systematic review // Therapeutic Advances in Medical Oncology. 2018. Vol. 10. 12 p. <https://doi.org/10.1177/1758835918786475>

3. Hoffmann C., Berganza C., Zhang J. Cold Atmospheric Plasma: methods of production and application in dentistry and oncology // Medical Gas Research. 2013. Vol. 3. Issue 1. P. 21. <https://doi.org/10.1186/2045-9912-3-21>

4. Jawaid P., Rehman M.U., Zhao Q.L., Takeda K., Ishikawa K., Hori M., et al. Helium-based cold atmospheric plasma-induced reactive oxygen species-mediated apoptotic pathway attenuated by platinum nanoparticles // Journal of Cellular and Molecular Medicine. 2016. Vol. 20. Issue 9. P. 1737–1748. <https://doi.org/10.1111/jcmm.12880>

5. Ermolaeva S.A., Varfolomeev A.F., Chernukha M.Yu., Yurov D.S., Vasiliev M.M., Kaminskaya A.A., et al. Bactericidal effects of non-thermal argon plasma in vitro, in biofilms and in the animal model of infected wounds // Journal of Medical Microbiology. 2011. Vol. 60. Pt. 1. P. 75–83. <https://doi.org/10.1099/jmm.0.020263-0>

6. Alkawareek M.Y., Gorman S.P., Graham W.G., Gilmore B.F. Potential cellular targets and antibacterial efficacy of atmospheric pressure non-thermal plasma // International Journal of Antimicrobial Agents. 2014. Vol. 43. Issue 2. P. 154–160. <https://doi.org/10.1016/j.ijantimicag.2013.08.022>

7. Flynn P.B., Busetti A., Wielogorska E., Chevallier O.P., Elliott C.T., Laverty G., et al. Non-thermal Plasma Exposure Rapidly Attenuates Bacterial AHL-Dependent Quorum Sensing and Virulence // Scientific Reports. 2016. Vol. 6. Article number: 26320. <https://doi.org/10.1038/srep26320>

8. Мартусевич А.К., Соловьева А.Г., Краснова С.Ю., Янин Д.В., Галка А.Г., Костров А.В.

Влияние гелиевой холодной плазмы на метаболические и физико-химические параметры крови человека *in vitro* // Биомедицина. 2018. N 2. C. 47–58.

9. Brun P., Pathak S., Castagliuolo I., Palù G., Zuin M., Cavazzana R., et al. Helium generated cold plasma finely regulates activation of human fibroblast-like primary cells // PLoS ONE. 2014. Vol. 9. Issue 8. P. e104397. <https://doi.org/10.1371/journal.pone.0104397>

10. Wiegand C., Fink S., Beier O., Horn K., Pfuch A., Schimanski A., et al. Dose- and time-dependent cellular effects of cold atmospheric pressure plasma evaluated in 3D skin models // Skin pharmacology and physiology. 2016. Vol. 29. Issue 5. P. 257–265. <https://doi.org/10.1159/000450889>

11. Мартусевич А.К., Соловьева А.Г., Янин Д.В., Галка А.Г., Краснова С.Ю. Влияние гелиевой холодной плазмы на параметры окислительного метаболизма крови *in vitro* // Вестник новых медицинских технологий. 2017. Т. 24. N 3. С. 163–166. https://doi.org/10.12737/article_59c4a9e679ca86.74880803

12. Мартусевич А.К., Соловьева А.Г., Краснова С.Ю. Влияние гелиевой холодной плазмы на состояние окислительного метаболизма крови крыс // Вестник Ульяновской государственной сельскохозяйственной академии. 2018. N 2 (42). С. 161–165. <https://doi.org/10.18286/1816-4501-2018-2-161-165>

13. Мартусевич А.К., Соловьева А.Г., Галка А.Г., Козлова Л.А., Янин Д.В. Влияние гелиевой холодной плазмы на метаболизм эритроцитов // Бюллетень экспериментальной биологии и медицины. 2019. Т. 167. N 2. С. 144–146.

14. Мартусевич А.К., Краснова С.Ю., Перетягин П.В., Галка А.Г., Голыгина Е.С., Костров А.В. Влияние гелиевой холодной плазмы на параметры вариабельности сердечного ритма крыс // Биофизика. 2019. Т. 64. N 3. С. 596–600. <https://doi.org/10.1134/S0006302919030219>

15. Мартусевич А.К., Краснова С.Ю., Галка А.Г., Перетягин П.В., Янин Д.В., Костров А.В. Оценка микроциркуляторного ответа на воздействие

холодной гелиевой плазмы // Биофизика. 2019. Т. 64. № 4. С. 767–771. <https://doi.org/10.1134/S0006302919040161>

16. Мартусевич А.К., Соловьева А.Г., Перетягин С.П. Влияние различных форм оксида азота на свойства альдегиддегидрогеназы эритроцитов // Вопросы биологической, медицинской и фармацевтической химии. 2014. № 11. С. 60–65.

17. Давыдюк А.В., Мартусевич А.К., Соловьева А.Г., Каримова Р.Г. Метаболическая адаптация эритроцитарных оксидоредуктаз к воз-

действию глутатион-содержащих динитрозильных комплексов железа // Ученые записки Казанской государственной академии ветеринарной медицины им. Н.Э. Баумана. 2015. Т. 221. № 1. С. 60–64.

18. Соловьева А.Г., Перетягин С.П. Влияние субхронического воздействия ингаляций оксида азота на метаболические процессы в крови экспериментальных животных // Биомедицинская химия. 2016. Т. 62. № 2. С. 212–214. <https://doi.org/10.18097/PBMC20166202212>

Contribution

Andrew K. Martusevich, Anna G. Soloveva, Svetlana Yu. Krasnova, Alexandr G. Galka, Alexandr V. Kostrov carried out the experimental work. The authors on the basis of the results summarized the material and wrote the manuscript. Andrew K. Martusevich, Anna G. Soloveva, Svetlana Yu. Krasnova, Alexandr G. Galka, Alexandr V. Kostrov have equal author's rights and bear equal responsibility for plagiarism.

Conflict interests

The authors declare no conflict of interests regarding the publication of this article.

The final manuscript has been read and approved by all the co-authors.

INFORMATION ABOUT THE AUTHORS

Andrew K. Martusevich,

Dr. Sci. (Biology), Head of Medical Biophysics Laboratory
Privolzhsky Research Medical University,
18/1 Verkhnevolzhskaya Emb.,
Nizhny Novgorod 603155,
Russian Federation,
✉ e-mail: cryst-mart@yandex.ru

Anna G. Soloveva,

Cand. Sci., (Biology), Head
of Experimental Medicine Laboratory,
Privolzhsky Research Medical University,
18/1 Verkhnevolzhskaya Emb.,
Nizhny Novgorod 603155,
Russian Federation
e-mail: cryst-mart@yandex.ru

Svetlana Yu. Krasnova,

Junior Scientist,
Medical Biophysics Laboratory,
Privolzhsky Research Medical University,
18/1 Verkhnevolzhskaya Emb.,
Nizhny Novgorod 603155,
Russian Federation,
e-mail: cryst-mart@yandex.ru

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СВЕДЕНИЯ ОБ АВТОРАХ

Мартусевич Андрей Кимович,

д.б.н., руководитель лаборатории
медицинской биофизики
Университетской клиники,
Приволжский исследовательский медицинский
университет Минздрава России,
603155, Нижний Новгород,
Верхневолжская наб., 18/1,
Российская Федерация,
✉ e-mail: cryst-mart@yandex.ru

Соловьева Анна Геннадьевна,

к.б.н., руководитель лаборатории
экспериментальной медицины
Университетской клиники,
Приволжский исследовательский медицинский
университет Минздрава России,
603155, Нижний Новгород,
Верхневолжская наб., 18/1,
Российская Федерация,
✉ e-mail: cryst-mart@yandex.ru

Краснова Светлана Юрьевна,

младший научный сотрудник
лаборатории медицинской биофизики
Университетской клиники,
Приволжский исследовательский медицинский
университет Минздрава России,
603155, Нижний Новгород,
Верхневолжская наб., 18/1,
Российская Федерация,
✉ e-mail: cryst-mart@yandex.ru

Alexandr G. Galka,
Junior Scientist,
Medical Biophysics Laboratory
Privolzhsky Research Medical University,
18/1 Verkhnevolzhskaya emb.,
Nizhny Novgorod 603155,
Russian Federation;
Junior Scientist,
Laboratory of Plasma Physics Modeling
Institute of Applied Physics RAS,
46 Ulyanov St., Nizhny Novgorod 603950,
Russian Federation,
e-mail: cryst-mart@yandex.ru

Галка Александр Георгиевич,
младший научный сотрудник лаборатории
моделирования космической плазмы
Университетской клиники,
Приволжский исследовательский медицинский
университет Минздрава России,
603155, Нижний Новгород,
Верхневолжская наб., 18/1,
Российская Федерация;
младший научный сотрудник лаборатории
медицинской биофизики,
Федеральный исследовательский центр
«Институт прикладной физики РАН»,
603950, г. Нижний Новгород, ул. Ульянова, 46,
Российская Федерация,
e-mail: cryst-mart@yandex.ru

Alexandr V. Kostrov,
Dr. Sci. (Physics and Mathematics), Professor,
Head of the Laboratory of Plasma Physics Modeling
Institute of Applied Physics of RAS,
46 Ulyanov St., Nizhny Novgorod 603950,
Russian Federation,
e-mail: cryst-mart@yandex.ru

Костров Александр Владимирович,
д.ф.-м.н., профессор, заведующий лабораторией
моделирования космической плазмы,
Федеральный исследовательский центр
«Институт прикладной физики РАН»,
603950, г. Нижний Новгород, ул. Ульянова, 46,
Российская Федерация,
e-mail: cryst-mart@yandex.ru